

(19)



Europäisches Patentamt
European Patent Office
Office européen des brevets



(11) Publication number:

0 510 678 A2

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: 92107043.9

(51) Int. Cl.⁵: **C12N 15/80, C12N 15/14,**
C12N 1/16, C12N 1/32,
/(C12N1/16,C12R1:84)

(22) Date of filing: 24.04.92

The applicant has filed a statement in accordance with Rule 28 (4) EPC (issue of a sample only to an expert). Accession number(s) of the deposit(s): NRRL B-18114, Y-15851, Y-18014, Y-18017, Y-11430 and Y-11431.

(30) Priority: 26.04.91 US 691079

(43) Date of publication of application:
28.10.92 Bulletin 92/44

(64) Designated Contracting States:
AT BE CH DE DK ES FR GB GR IT LI LU NL PT SE

(71) Applicant: **PHILLIPS PETROLEUM COMPANY**
5th and Keeler

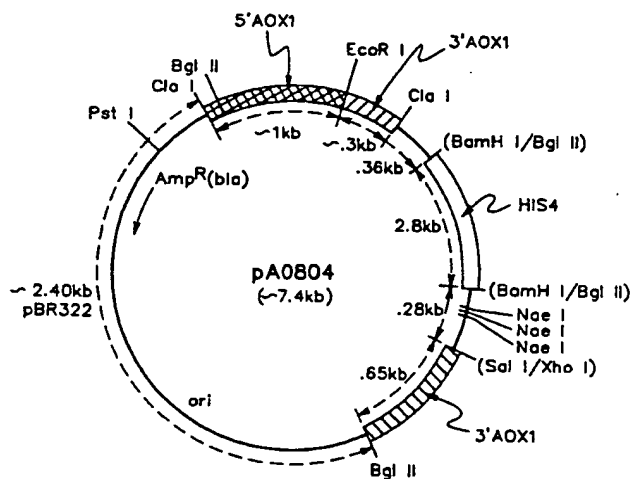
Bartlesville Oklahoma 74004(US)

(72) Inventor: **Prevatt, William Dudley**
4524 Barlow Drive
Bartlesville, OK 74006(US)
Inventor: **Sreekrishna, Kotikanyad**
1060 Rolling Meadow Court
Bartlesville, OK 74006(US)

(74) Representative: **Dost, Wolfgang,**
Dr.rer.nat.,Dipl.-Chem. et al
Patent- & Rechtsanwälte Bardehle .
Pagenberg . Dost . Altenburg . Frohwitter .
Geissler & Partner Galileiplatz 1 Postfach 86
06 20
W-8000 München 86(DE)

(54) Expression of human serum albumin in *pichia pastoris*.

(57) A process for the production of HSA in *Pichia pastoris* cells comprising cultivating *Pichia pastoris* cells capable of expressing HSA at a pH of about 5.7 to about 6.4 contemporaneously with the expression of HSA.

**FIG. 1**

Field of the Invention

This invention relates to the field of recombinant DNA biotechnology. In one aspect, this invention relates to a process for the improved expression of secreted human serum albumin (HSA) in *Pichia pastoris*.

Background

Human serum albumin is the most abundant plasma protein of adults. The concentration of albumin is 40 mg/ml, or 160g of albumin circulating throughout the human body for a 70 Kg adult male. This protein maintains osmotic pressure and functions in the binding and transport of copper, nickel, calcium (weakly, at 2-3 binding sites), bilirubin and protoporphyrin, long-chain fatty acids, prostaglandins, steroid hormones (weak binding with these hormones promotes their transfer across the membranes), thyroxine, triiodothyronine, cystine, and glutathione. According to Peters, T. and Reed, R. G. in *Albumin: Structure, Biosynthesis and Function*, (Peter, T. and Sjöholm, J. eds.) 1977 p.11-20, over 10,000 kilograms of purified albumin are administered annually in the United States alone to patients with circulatory failure or with albumin depletion.

Currently the only commercial source of HSA is from fractionated blood. Considering the possible dangers of blood borne contaminants and pathogens, it would be a considerable contribution to the commercial production of HSA to develop alternate methods of producing HSA. With the advent of recombinant DNA technology, it is now possible to produce HSA by alternate methods.

HSA has also been expressed in *Saccharomyces cerevisiae* as disclosed by Etcheverry et al. in *Bio/technology*, August 1986, p. 726 and Arjum Singh in EPA 123,544. Etcheverry disclosed HSA expression intracellularly in a concentration of approximately 6 mg/l and the secretion of HSA which remained cell associated. Arjum Singh also disclosed the expression of HSA in *Saccharomyces cerevisiae* in combination with the α -factor promoter and signal sequence. Singh appears to have been able to achieve an intracellular production level of approximately 25 mg/l and a secreted production level of 3 mg/l. *Pichia pastoris* has also been used to express HSA as is disclosed in EPA 344,459. The concentration of HSA produced in *Pichia pastoris* appears to be about 89 ng HSA/mg of protein. Although the process for producing HSA in recombinant expression systems has been established by these experiments it would be desirable to optimize these processes to achieve the maximum possible HSA production.

Therefore, it would be a significant contribution to the art to provide a process for increasing the yield of HSA from the recombinant expression of HSA in microorganism such as *Pichia pastoris*.

Therefore, it is an object of this invention to provide a process for increasing the yield of HSA produced in a recombinant expression systems.

Summary of the Invention

In accordance, we have discovered a process for improving the secreted expression of HSA in *Pichia pastoris* cells comprising:

(a) cultivating in a fermentation broth transformed *Pichia pastoris* cells capable of expressing HSA under conditions suitable for the sustained viability of said *Pichia pastoris* cells under suitable conditions for the expression of HSA by said *Pichia pastoris* cells; and maintaining the pH of said fermentation broth from a pH of from about 5.7 to about 6.0 contemporaneously with the expression of HSA.

Detailed Description of the Figures

Figure 1 provides a representation of plasmid pAO804 which contains a linear site-specific integrative vector in the fragment clockwise from BglII to BglII. The structural gene may be inserted in the unique EcoRI site of this plasmid. This plasmid may be recovered from the plasmid DNA of NRRL B-18114 by EcoRI digest and gel electrophoresis to recover a linear ~7.4 kb EcoRI fragment corresponding to Figure 1.

Figure 2 provides a representation of pHSA13 in circular form.

Figure 3 provides a restriction map of the AOX1 5' regulatory region isolated from *Pichia pastoris*.

Figure 4 provides a restriction map of the DAS1 5' regulatory region isolated from *Pichia pastoris*.

Figure 5 provides a restriction map of the AOX1 3' termination sequence isolated from *Pichia pastoris*.

Figure 6 provides a restriction map of the DAS1 3' termination sequence isolated from *Pichia pastoris*.

Figure 7 provides a representation of pHSA113 in linear form.

Figure 8 provides a representation of plasmid pAO807N which contains a linear site-specific integrative

vector in the fragment clockwise from NotI to NotI. The structural gene may be inserted in the unique EcoRI site of this plasmid.

Detailed Description

5

Generally *Pichia pastoris* is optimally grown at from about pH 4.8 to about pH 5.2. Between this pH range *Pichia pastoris* provided with a suitable nutrient media exhibits robust growth. This pH range also appears to result in high levels of expression of several foreign proteins such as hepatitis B surface antigen. This pH range also appeared to provide high levels of expression with human serum albumin (HSA). For example growing *Pichia pastoris* cells which had been transformed with a vector containing a HSA structural gene operably linked to a 5' regulatory region (i.e. a promoter) and a 3' termination sequence, the expression levels of HSA which had been obtained were approximately .71 to .81 grams/liters of HSA in the fermentation broth. However, we have been able to further increase this yield by at least 50% by taking the unprecedented step of shifting the pH of the fermentation broth from about 5.2 to in the range of from about pH 5.7 to about pH 6.4, with a preferred pH range of from about pH 5.7 to about pH 6.0 and most preferably a pH in the range of from pH 5.75 to pH 5.85. The increased secretion levels obtained in the upper limits of the pH range (i.e. from in the range of pH 6.0 to pH 6.4) have been confirmed in shake tube optimization studies which indicate that the presence of yeast extract and peptone together with aeration will provide optimal HSA secretion in shake tubes. However, the use of yeast extract, peptone and excess aeration is not believed necessary in large scale fermentation where the pH can be continuously monitored. We believe that this higher pH level will increase the yield of any *Pichia pastoris* strain transformed with an expression cassette containing a promoter and a structural gene encoding a signal sequence and the mature HSA protein. Further it would appear that this result will be applicable to a variety of heterologous structural genes which encode a signal sequence and a mature heterologous protein. Suitable heterologous proteins which may be expressed at higher levels utilizing this method include but are not limited to heterologous proteins selected from the group consisting of tissue plasminogen activator, albumins (such as human serum albumin), lysozymes (such as bovine lysozyme), interferons (such as gamma-interferon and beta-interferon) and invertase. Each of the heterologous structural genes utilized in the present invention must have a signal sequence operably linked to the 5' end of sequence coding for the mature heterologous protein to effect the secretion of the mature protein. For example the tissue plasminogen activator, human serum albumins, bovine lysozyme, beta-interferon, gamma-interferon and invertase proteins may all be secreted utilizing the native signal sequence. Furthermore these proteins may also be secreted utilizing secretion signal sequences from *Pichia pastoris* such as the acid phosphatase signal sequence disclosed in U.S. Patent Application Serial Number 07/627,539 filed December 14, 1990 by Richard Buckholz assigned to Phillips Petroleum Company (incorporated herein by reference) or the alpha-mating factor signal sequence from *Saccharomyces cerevisiae*.

Utilizing the present invention, HSA secretion levels of approximately 1-3 grams of authentic HSA per liter of fermentation broth have been obtained. This invention thus provides a means for the high level secretion of HSA. Achieving these levels of HSA production is a significant advancement over the prior production levels, since at the level of 1-3 grams per liter the recovery of HSA in high yields with high purities is possible.

To express the HSA structural gene, the gene must be operably linked to a 5' regulatory region and a 3' termination sequence, which forms an expression cassette which will be inserted into a host (usually a microorganism) via a vector (such as a plasmid or linear site-specific integrative vector). Operably linked as used in this context refers to a juxtaposition wherein the 5' regulatory region, structural gene, and 3' termination sequence are linked and configured so as to perform their normal function. 5' regulatory region or promoter as used herein means DNA sequences which respond to various stimuli and provide enhanced rates of mRNA transcription. 3' termination sequence are sequences 3' to the stop codon of a structural gene which function to stabilize the mRNA transcription product of the gene to which the sequence is operably linked (such as sequences which elicit polyadenylation). For the practice of this invention, it is preferred that the ATG of the structural gene be linked with as few intervening deoxyribonucleotides as possible to the 3' end of the 5' regulatory region, preferably about 11 or less deoxyribonucleotides and most preferably 8 or less deoxyribonucleotides. It is also preferred that the adenine and thymine content of the intervening deoxyribonucleotides be in the range of from about 55 percent to about 64 percent. Further, it appears that there are nucleotide preferences for certain specific locations. Counting left from the ATG codon of the structural gene with the first position left being the -1 position, it appears that adenine or cytosine is the most preferred deoxyribonucleotide, in the -2 position the most preferred deoxyribonucleotide is either adenine or thymine, in the -3 position the most preferred deoxyribonucleotide is

adenine or thymine and the most preferred nucleotide at the -4 position is adenine, thymine or cytosine. Currently, it is preferred that the AOX1 or DAS1 5' regulatory region having the restriction maps of Figures 3 and 4 or, the sequences provided as SEQ ID No: 1 and SEQ ID No: 2, respectively, be linked at their 3' end of the sequence to the ATG start codon of the HSA structural gene. One example of an appropriate linkages for the AOX1 5' regulatory region is illustrated below:

Table I

Construct Designation	End of the 5' Regulatory Region for <u>AOX 1</u>	Deoxyribonucleotide intervening before ATG start condon
pHSA413	5' - TTCGAAACG	5' - NONE

Several 5' regulatory regions have been characterized and can be employed in conjunction with the expression of HSA in *Pichia pastoris*. Exemplary 5' regulatory regions are the primary alcohol oxidase (AOX1), dihydroxyacetone synthase (DAS1), glyceraldehyde-3-phosphate dehydrogenase gene (GAP), acid phosphatase gene (PHO1) and the p40 regulatory regions, derived from *Pichia pastoris* and the like. The AOX1 5' regulatory region, DAS1 5' regulatory region and p40 5' regulatory region are described in U.S. Patent 4,855,231, incorporated herein by reference. The GAP 5' regulatory region is disclosed in EPA 374,913 published June 27, 1990, incorporated herein by reference. The PHO1 5' regulatory region is disclosed in U.S. Patent Application 07/672,539 filed December 14, 1990, assigned to Phillips Petroleum Company. The presently preferred 5' regulatory regions employed in the practice of this invention are those characterized by their ability to respond to methanol-containing media, such regulatory regions selected from the group consisting of AOX1, and DAS1. The most preferred 5' regulatory region for the practice of this invention is the AOX1 5' regulatory region.

3' termination sequences should be utilized in the expression cassette as discussed above. 3' termination sequences may function to terminate, polyadenylate and/or stabilize the messenger RNA coded for by the structural gene when operably linked to a gene, but the particular 3' termination sequence is not believed to be critical to the practice of the present invention. A few examples of illustrative sources for 3' termination sequences for the practice of this invention include but are not limited to the *Hansenula polymorpha* and *Pichia pastoris* 3' termination sequences. Preferred are those derived from *Pichia pastoris* such as those selected from the group consisting of the 3' termination sequences of AOX1 gene, DAS1 gene, p40 gene, GAP gene, PHO1 gene and HIS4 gene. Particularly preferred is the 3' termination sequence of the AOX1 gene.

Pichia pastoris may be transformed with a variety of HSA structural genes (in the inventive transformants discussed herein the HSA structural gene encodes both a signal sequence and a mature HSA protein). HSA structural genes have been sequenced by Lawn et al. *Nuc. Acids Res.* 9:6105 (1981), and Dugaiczky et al., *Proc. Natl. Acad. Sci. USA* 79:71 (1982). These genes may also be obtained by reisolation of the genes by the technique of Lawn et al., Dugaiczky et al. or synthesized *in vitro* by a custom gene manufacturer such as British Biotechnology, Ltd. One possible method of obtaining a HSA gene would be to screen a human liver cDNA library with oligonucleotide probes or screen a human liver cDNA expression library with anti-HSA antisera to identify HSA expressing cDNAs. One suitable HSA structural gene is provided in SEQ ID NO: 3. Once a structural gene for HSA is recovered, it may be necessary to further tailor the gene. Following the isolation of an HSA structural gene, the gene is inserted into a suitable *Pichia pastoris* vector such as a plasmid or linear site-specific integrative vector.

Plasmid-type vectors have long been one of the basic elements employed in recombinant DNA technology. Plasmids are circular extra-chromosomal double-stranded DNA found in microorganisms. Plasmids have been found to occur in single or multiple copies per cell. Included in plasmid DNA is the information required for plasmid reproduction, e.g. an autonomous replication sequence such as those disclosed by James M. Cregg in U.S. Patent 4,837,148, issued June 6, 1989, incorporated herein by reference. Additionally one or more means of phenotypically selecting the plasmid in transformed cells may also be included in the information encoded in the plasmid.

Suitable integrative vectors for the practice of the present invention are the linear site-specific integrative vectors described by James M. Cregg, in U.S. Patent 4,882,279, issued November 21, 1989, which is incorporated herein by reference. These vectors comprise a serially arranged sequence of at least 1) a first insertable DNA fragment; 2) a selectable marker gene; and 3) a second insertable DNA fragment. An expression cassette containing a heterologous structural gene is inserted in this vector between the first and second insertable DNA fragments either before or after the marker gene. Alternatively, an expression

cassette can be formed *in situ* if a regulatory region or promoter is contained within one of the insertable fragments to which the structural gene may be operably linked.

The first and second insertable DNA fragments are each at least about 200 nucleotides in length and have nucleotide sequences which are homologous to portions of the genomic DNA of the species to be transformed. The various components of the integrative vector are serially arranged forming a linear fragment of DNA such that the expression cassette and the selectable marker gene are positioned between the 3' end of the first insertable DNA fragment and the 5' end of the second insertable DNA fragment. The first and second insertable DNA fragments are oriented with respect to one another in the serially arranged linear fragment as they are oriented in the parent genome.

Nucleotide sequences useful as the first and second insertable DNA fragments are nucleotide sequences which are homologous with separate portions of the native genomic site at which genomic modification is to occur. For example, if genomic modification is to occur at the locus of the alcohol oxidase gene, the first and second insertable DNA fragments employed would be homologous to separate portions of the alcohol oxidase gene locus. Examples of nucleotide sequences which could be used as first and second insertable DNA fragments are deoxyribonucleotide sequences selected from the group consisting of the *Pichia pastoris* alcohol oxidase (AOX1) gene, dihydroxyacetone synthase (DAS1) gene, p40 gene, glyceraldehyde-3-phosphate dehydrogenase (GAP), acid phosphatase (PHO1) and HIS4 gene. The AOX1 gene, DAS1 gene, p40 gene and HIS4 genes are disclosed in U.S. Patents 4,855,231 and 4,885,242 both incorporated herein by reference. The designation DAS1 is equivalent to the DAS designation originally used in U.S. Patents 4,855,231 and 4,885,242. The GAP gene is disclosed in EPA 374,913 published June 27, 1990 incorporated herein by reference. The PHO1 gene is disclosed in U.S. Patent Application 07/672,539 filed December 14, 1990, assigned to Phillips Petroleum Company, incorporated herein by reference.

The first insertable DNA fragment may contain an operable regulatory region which may comprise the regulatory region utilized in the expression cassette. The use of the first insertable DNA fragment as the regulatory region for an expression cassette is a preferred embodiment of this invention. Figure 1 provides a diagram of a vector utilizing the first insertable DNA fragment as a regulatory region for a cassette. Optionally, as shown in Figure 1, an insertion site or sites and a 3' termination sequence may be placed immediately 3' to the first insertable DNA fragment. This conformation of the linear site-specific integrative vector has the additional advantage of providing a ready site for insertion of a structural gene without necessitating the separate addition of a compatible 3' termination sequence.

If the first insertable DNA fragment does not contain a regulatory region, a suitable regulatory region will need to be inserted linked to the structural gene, in order to provide an operable expression cassette. Similarly, if no 3' termination sequence is provided at the insertion site to complete the expression cassette, a 3' termination sequence can be operably linked to the 3' end of the structural gene.

It is also highly desirable to include at least one selectable marker gene in the DNA used to transform the host strain. This facilitates selection and isolation of those organisms which have incorporated the transforming DNA. The marker gene confers a phenotypic trait to the transformed organism which the host did not have, e.g. restoration of the ability to produce a specific amino acid where the untransformed host strain has a defect in the specific amino acid biosynthetic pathway, or provides resistance to antibiotics and the like. Exemplary selectable marker genes may be selected from the group consisting of the HIS4 gene (disclosed in U.S. Patent 4,885,242) and the ARG4 gene (disclosed in U.S. Patent 4,818,700 incorporated herein by reference) from *Pichia pastoris* and *Saccharomyces cerevisiae*, the invertase gene (SUC2) (disclosed in U.S. Patent 4,857,467 incorporated herein by reference) from *Saccharomyces cerevisiae*, or the G418^R/kanamycin resistance gene from the *E. coli* transposable elements Tn601 or Tn903.

Those skilled in the art recognize that additional DNA sequences can also be incorporated into the vectors employed in the practice of the present invention, such as, for example, bacterial plasmid DNA, bacteriophage DNA, and the like. Such sequences enable the amplification and maintenance of these vectors in bacterial hosts.

The insertion of the HSA structural gene into suitable vectors may be accomplished by any suitable technique which cleaves the chosen vector at an appropriate site or sites and results in at least one operable expression cassette containing the HSA structural gene being present in the vector. Ligation of the HSA structural gene may be accomplished by any appropriate ligation technique such as utilizing T4 DNA ligase.

The initial selection, propagation, and optional amplification of the ligation mixture of the HSA structural gene and a vector is preferably performed by transforming the mixture into a bacterial host such as *E. coli* - (although the ligation mixture could be transformed directly into a yeast host but, the transformation rate would be extremely low). Suitable transformation techniques for *E. coli* are well known in the art.

Additionally, selection markers and bacterial origins of replication necessary for the maintenance of a vector in a bacterial host are also well known in the art. The isolation and/or purification of the desired plasmid containing the HSA structural gene in an expression system may be accomplished by any suitable means for the separation of plasmid DNA from the host DNA. Similarly the vectors formed by ligation may be tested, preferably after propagation, to verify the presence of the HSA gene and its operable linkage to a regulatory region and a 3' termination sequence. This may be accomplished by a variety of techniques including but not limited to endonuclease digestion, gel electrophoresis, or Southern hybridization.

Transformation of plasmids or linear vectors into yeast hosts may be accomplished by suitable transformation techniques including but not limited to those taught by Cregg and Barringer, U.S. Patent 4,929,555; Hinnen et al., *Proc. Natl. Acad. Sci.* 75, (1978) 1929; Ito et al., *J. Bacteriol.* 153, (1983) 163; Cregg et al. *Mol. Cell Biol.* 5 (1985), pg. 3376; D. W. Stroman et al., U.S. Patent 4,879,231, issued November 7, 1989; or Sreekrishna et al., *Gene*, 59 (1987), pg. 115. Preferable for the practice of this invention is the transformation technique of Cregg et al. (1985). It is desirable for the practice of this invention to utilize an excess of linear vectors and select for multiple insertions by Southern hybridization.

The yeast host for transformation may be any suitable methylotrophic yeast. Suitable methylotrophic yeasts include but are not limited to yeast capable of growth on methanol selected from the group consisting of the genera *Hansenula* and *Pichia*. A list of specific species which are exemplary of this class of yeasts may be found in C. Anthony, *The Biochemistry of Methylotrophs*, 269 (1982). Presently preferred are methylotrophic yeasts of the genus *Pichia* such as the auxotrophic *Pichia pastoris* GS115 (NRRL Y-15851); *Pichia pastoris* GS190 (NRRL Y-18014) disclosed in U.S. Patent 4,818,700; and *Pichia pastoris* PPF1 (NRRL Y-18017) disclosed in U.S. Patent 4,812,405. Auxotrophic *Pichia pastoris* strains are also advantageous to the practice of this invention for their ease of selection. It is recognized that wild type *Pichia pastoris* strains (such as NRRL Y-11430 and NRRL Y-11431) may be employed with equal success if a suitable transforming marker gene is selected, such as the use of SUC2 to transform *Pichia pastoris* to a strain capable of growth on sucrose or an antibiotic resistance marker is employed, such as G418.

Transformed *Pichia pastoris* cells can be selected for by using appropriate techniques including but not limited to culturing previously auxotrophic cells after transformation in the absence of the biochemical product required (due to the cell's auxotrophy), selection for and detection of a new phenotype ("methanol slow"), or culturing in the presence of an antibiotic which is toxic to the yeast in the absence of a resistance gene contained in the transformant.

Isolated transformed *Pichia pastoris* cells are cultured by appropriate fermentation techniques such as shake flask fermentation, high density fermentation or the technique disclosed by Cregg et al. in, *High-Level Expression and Efficient Assembly of Hepatitis B Surface Antigen in the Methylotrophic Yeast, Pichia Pastoris* 5 Bio/Technology 479 (1987). Isolates may be screened by assaying for HSA production to identify those isolates with the highest HSA production level.

The cultivation of transformed *Pichia pastoris* can be conducted in an aqueous continuous or batch-fed manner, utilizing a variety of carbon-energy sources and/or nutrient sources. For the practice of the present invention, batch-fed fermentation is preferred. Suitable carbon-energy sources for growing *Pichia pastoris* include but are not limited to the carbon-energy source selected from the group consisting of methanol, glycerol, sorbitol, glucose, fructose and combinations of any two or more thereof. Preferred carbon-energy sources for growing *Pichia pastoris* are carbon-energy sources selected from the group consisting of methanol, glycerol, and combinations thereof. A suitable nutrient source or media for *Pichia pastoris* would include at least one nitrogen source, at least one phosphate source, at least one source of minerals such as iron, copper, zinc, magnesium, manganese, calcium, and other trace elements, and vitamins (such as biotin, pantothenic acid, and thiamine as required).

Suitable sources of at least one carbon-energy source and nutrients can be obtained from a variety of sources or may consist of a single source. However, preferred are at least one carbon-energy source and/or nutrient sources which have a defined character. One carbon-energy source and/or nutrient composition which has proven effective is:

Table II

Carbon-Energy Source and Nutrients	
Component per Liter of Water	
Carbon-energy Source (glycerol)	50.0 g/l
H ₃ PO ₄ (85%)	21 ml/l
CaSO ₄ · 2H ₂ O	0.9 g/l
K ₂ SO ₄	14.28 g/l
MgSO ₄ · 7H ₂ O	11.7 g/l
KOH	3.9 g/l
Peptone	10.0 g/l
¹ Yeast Extracts	5.0 g/l
² Minerals and Trace Metals	1.0 ml/l

¹Yeast extract is Amberex™ 1003 which is available from and a trademark of Universal Foods Corporation, Milwaukee, Wisconsin.

²Minerals and trace metals are FeSO₄ · 7H₂O 65.0 g/l, CuSO₄ · 5H₂O 6.0 g/l, ZnSO₄ · 7H₂O 20 g/l, MnSO₄ 3.0 g/l and H₂SO₄ 5.0 ml/l

The yeast extracts utilized in the present invention include but are not limited to yeast extracts selected from the group consisting of Amberex™ 1003 and Bacto™ Yeast Extract (Difco Laboratories Incorporated). Alternatively, corn steep liquor could be used to replace yeast extracts as a source of nitrogen.

Trace metals utilized in the present invention are those trace metals generally utilized in the yeast growth provided in an amount sufficient to not limit the growth rate or HSA production of *Pichia pastoris* which include but are not limited to trace metals selected from the group consisting of cobalt, molybdenum, iron, copper, zinc, and manganese.

The fermentation temperature should generally range from about 20°C to about 35°C and preferably should be about 30°C.

The dissolved oxygen content in the fermentation vessel where the fermentation is conducted in a batch-fed manner may range from about 20 percent to about 80 percent of saturation and preferably will range from about 30 percent to about 60 percent of saturation.

After the *Pichia pastoris* strains transformed with a vector containing the HSA structural gene have been cultivated to a high density, the transformed strains should then be induced to express HSA at a pH of from about 5.7 to about 6.0. For example, if this technique is employed with a strain transformed with a linear expression cassette containing a methanol inducible regulatory region, the culture would first be grown to the desired density on minimal salts, biotin and 5 percent glycerol by weight. The pH should be adjusted to 5.8 (with ammonia) with a temperature of about 30°C and a dissolved oxygen concentration of about 20 percent of saturation. After the glycerol is exhausted, the promoter would be induced by beginning a slow methanol feed. The feed should provide methanol to the culture at a rate at least sufficient to maintain the viability of the culture but the maximum methanol concentration in contact with the culture should be no more than about 5.0 percent by weight. The HSA secretion can be monitored during the methanol feeding by sampling the HSA present in the cell free broth. Suitable test for quantifying the amount of HSA produced are known to those skilled in the art, such as running polyacrylamide gels. The methanol feed should be continued until the HSA concentration reaches an acceptable level. Generally, the HSA production will peak after about 120 hours on methanol feed.

If the transformed *Pichia pastoris* cells are grown in shake tubes or shake flasks instead of pH controlled fermenter, additional steps should be taken to assure the maximum yields of secreted proteins, such as HSA. Specifically, it is recommended that the media used be modified from that used in fermenter to a complex media and the aeration be increased. The complex media utilized in the shake flasks and shake tubes should contain added amino acids. The amino acids may be in a defined media containing glutamic acid, methionine, lysine, leucine, isoleucine and other amino acids or through a complex media supplement, such as yeast extract or casamino acids. The relative concentrations of the added amino acids should generally range from about 2.5 mg/liter to about 10 mg/liter with the preferred range being from about 4 mg/liter to about 6 mg/liter of glutamic acid, methionine, lysine, leucine and isoleucine and from about 0.5 mg/liter to about 3 mg/liter of the remaining amino acid (however, histidine may be omitted entirely from the added amino acids). If yeast extract is used in place of the added amino acids, it is preferred that the yeast extract be provided in a concentration of in the range of from about 1 g/liter to

about 15 g/liter be utilized in the media and most preferably the yeast extract will be provided in a concentration of 10 g/liter. It has also been found desirable to add peptone to the media to improve secretion in shake tubes and shake flasks. For optimum secretion that peptone be used with the yeast extract in a concentration of from in the range of from about 1 g/liter to about 50 g/liter, and most preferably in a concentration of about 20 g/liter. As a guideline, it is generally recommended that the peptone concentration be twice the yeast extract concentration.

Aeration in shake flask and shake tube growth of transformed *Pichia pastoris* appears to be an important parameter in obtaining optimum secretion. To insure adequate aeration, it is recommended that shake tube or flask have a large aperture covered with an air permeable cap. Suitable air permeable caps can be made of a loose filter material, such as cheese cloth. One suitable shake flask for this invention is the Tunair shake flask. Generally, low baffle shake flasks are also recommended to avoid excessive foaming. Shaker speed for aeration is recommended to be in the range of from about 250 rpms to about 300 rpms.

After a suitable cell density is achieved in the shake flask or shake tube, the cells may be recovered then resuspended in a medium containing methanol in place of the carbon source used for growth to induce the secretion of protein. The flask or shake tubes may then be monitored on a regular basis to determine when the desired level of production has been achieved.

The invention will now be described in greater detail in the following non-limiting examples.

Examples

General information pertinent to the Examples:

Strains

Pichia pastoris GS115 (his 4) NRRL Y-15851
E. coli DG75' (hsd1, leu6, lacY, thr-1, supE44, tonA21, lambda⁻)

Buffers, Solutions and Media

The buffers, solutions, and media employed in the following examples have the compositions given below:

dH₂O

deionized H₂O that has been treated with a milli-Q (Millipore) reagent water system.

1M Tris buffer

121.1 g Tris base in 800 mL of H₂O; adjust pH to the desired value by adding concentrated (35%) aqueous HCl; allow solution to cool to room temperature before final pH adjustment, dilute to a final volume of 1 L.

TE buffer

1.0 mM EDTA
 in 0.01 M (pH 8.0) Tris buffer

SED

1 M sorbitol
 25 mM EDTA
 50 mM DTT, added prior to use
 --adjust to pH 8

SCE

9.1 g sorbitol
 1.47 g Sodium citrate
 0.168 g EDTA
 --pH to 5.8 with HCl in 50 ml
 dH₂O and autoclave

CaS

1 M sorbitol
 10 mM CaCl₂
 --filter sterilize

SOS:

1 M sorbitol

PEG

0.3x YPD
 10 mM CaCl₂
 20% polyethylene glycol-3350
 10 mM CaCl₂

		10 mM Tris-HCl (pH 7.4)
		--filter sterilize
	Solution A	0.2 M Tris-HCl (pH 7.5)
5		0.1 M MgCl ₂
		0.5 M NaCl
		0.01 M dithiothreitol (DTT)
	Solution B	0.2 M Tris-HCl (pH 7.5)
		0.1 M MgCl ₂
		0.1 M DTT
10	Solution C (keep on ice)	4 µl solution B
		4 µl 10 mM dATP
		4 µl 10 mM dTTP
		4 µl 10 mM dGTP
		4 µl 10 mM dCTP
15		4 µl 10 mM ATP
		5 µl T ₄ ligase (2 U/µl)
		12 µl H ₂ O
		Recipe for Solution C was modified from Zoller & Smith
20	LB Broth, 1 liter	5.0 g yeast extract
		10.0 g tryptone
		5.0 g NaCl
	10X Transfer Buffer	96.8 g Trizma Base
		9.74 g glycine
25		water to 1 liter
	Ligation Buffer	50 mM Tris-HCl (pH 7.4)
		10 mM MgCl ₂
		10 mM dithiothreitol
		1 mM ATP
30	Phosphatase Buffer	50 mM Tris-HCl (pH 9.0)
		1 mM MgCl ₂
		1 mM ZnCl ₂
		1 mM spermidine
	<u>Bsu36I</u> buffer	100 mM NaCl
35		10 mM Tris-HCl (pH 7.4)
		10 mM MgCl ₂
		100 µg/ml BSA
	<u>Csp45I</u> buffer	60 mM NaCl
		10 mM Tris-HCl, pH 7.5
40		7 mM MgCl ₂
		100 µg/ml BSA
	REact 1 buffer	50 mM Tris-HCl, pH 8.0
		10 mM MgCl ₂
		100 µg/ml BSA
45	REact 2 buffer	REact 1 buffer + 50 mM NaCl
	REact 3 buffer	REact 1 buffer + 100 mM NaCl
	HS buffer	50 mM Tris-HCl, pH 7.5
		10 mM MgCl ₂
		100 mM NaCl
50		1 mM DTT
		100 µg/ml BSA
	10X Basal Salts	42 mls Phosphoric Acid, 85%
		1.8 g Calcium Sulfate • 2H ₂ O
		28.6 g Potassium Sulfate
55		23.4 g Magnesium Sulfate • 7H ₂ O
		6.5 g Potassium Hydroxide
	Ptm ₁ Trace Salts Solution	6.0 g Cupric Sulfate • 5H ₂ O
		0.08 g Sodium Iodide

	3.0 g Manganese Sulfate • H ₂ O
	0.2 g Sodium Molybdate • H ₂ O
	0.02 g Boric Acid
5	0.5 g Cobalt Chloride
	20.0 g Zinc Chloride
	65.0 g Ferrous Sulfate • H ₂ O
	0.20 g Biotin
	5.0 mls Sulfuric Acid
10	YPD (yeast extract peptone dextrose medium)
	10 g bacto yeast extract
	20 g peptone
	10 g dextrose
	water to 1 liter
	MGY (minimal glycerol medium)
15	13.4 g yeast nitrogen base with ammonium sulfate, and without amino acids
	400 µg biotin
	10 ml glycerol
	water to 1 liter
	MM (minimal methanol medium)
	Same as MGY, except that 5 ml methanol is used in the place of 10 ml glycerol.
20	SDR (supplemented dextrose regeneration medium):
	13.4 g yeast nitrogen base with ammonium sulfate and without amino acids
	400 µg biotin
	182 g sorbitol
25	10 g glucose
	2 g Histidine assay mix (Gibco)
	50 mg glutamine
	50 mg methionine
	50 mg lysine
30	50 mg leucine
	50 mg isoleucine
	10 g agarose
	water to 1 liter
	BMGR (Buffered minimal glycerol-enriched medium)
35	100 ml/liter Potassium phosphate buffer, (pH 6.0)
	13.4 grams/liter Yeast nitrogen base with ammonium sulfate
	400 µg/liter biotin
	10 ml/liter glycerol
40	Amino acids
	glutamic acid, methionine, lysine, leucine and isoleucine: each at 5 mg/liter;
	all the other amino acids except histidine at 1 mg/liter
45	Nucleotides
	adenine sulfate, guanine hydrochloride, uracil, and xanthine, each at 40 µg/liter
	Vitamins
	thiamine hydrochloride, riboflavin, and calcium pantothenate, each at 2 µg/liter;
50	pyridoxide hydrochloride and nicotinic acid, each at 4 µg/liter;
	pyridoxamine hydrochloride and pyridoxal hydrochloride, each at 1 µg/liter;
	para-amino benzoic acid at 0.3 µg/liter;
55	folic acid at 0.03 µg/liter
	Trace minerals
	magnesium sulfate at 800 µg/liter;
	ferrous sulfate at 40 µg/liter;

	manganese sulfate at 80 µg/liter; sodium chloride at 40 µg/liter
BMGY (Buffered minimal glycerol-complex medium)	100 ml/liter potassium phosphate buffer, (pH 6.0)
5	13.4 grams/liter yeast nitrogen base with ammonium sulfate and without amino acids
	biotin at 400 µg/liter
	glycerol at 10 ml/liter
	yeast extract at 10 g/liter
10	peptone at 20 g/liter
BMMR (Buffered minimal methanol-enriched medium)	Same as BMGR, with the exception that 5 ml methanol/liter is added in the place of glycerol
BMMY (Buffered minimal methanol-complex medium)	Same as BMGY, with the exception that 5 ml methanol/liter is added in the place of glycerol

15 Techniques

Suitable techniques for recombinant DNA lab work may be found in many different references including but not limited to: Methods in Enzymology, (Orlando, FL: Academic Press, Inc.), particularly Volume 152, published as, Guide to Molecular Cloning Techniques, by Berger and Kimmel (Orlando, FL: Academic Press, Inc., 1987) and Molecular Cloning/A Laboratory Manual, by Sambrook et al., 2d ed. (Cold Spring Harbor Laboratory Press, 1989) and which are all hereby incorporated by reference.

20 Example I

25 Construction of 5'-exact HSA expression vector pHSA313

The pHSA313 vector was constructed to provide a vector with an exact linkage between the 3' end of the native AOX1 5' regulatory region (promoter) and the start codon of the HSA structural gene.

30 A. Creation of pHSA113ΔCla

About 200 ng of pHSA113, disclosed in European Patent Application 0 344 459 which is herein incorporated by reference, (see Figure 7) was digested at 37° C for 1 hour with 1 unit of ClaI in 20 µl of REact 1 buffer. The digestion mixture was brought to 100 µl with water and extracted once with an equal volume of phenol:chloroform:isoamyl alcohol (25:24:1 V/V), followed by extracting the aqueous layer with an equal volume of chloroform:isoamyl alcohol (24:1). The DNA in the aqueous phase was precipitated by adjusting the NaCl concentration to 0.2 M and adding 3 volumes of cold ethanol. The mixture was allowed to stand on ice (4° C) for 10 minutes and the DNA precipitate was collected by centrifugation for 30 minutes at 10,000 x g in a microfuge at 4° C. The DNA pellet was washed 2 times with 70% aqueous cold ethanol. The washed pellet was vacuum dried and dissolved in 10 µl water to which 2 µl of 10 x ligation buffer, 2 µl of 1 mg/ml BSA, 6 µl of water and 1 unit T₄ DNA ligase were added. The mixture was incubated overnight at 4° C and a 10 µl aliquot was used to transform E. coli DG75' (Maniatis, et al.) to obtain pHSA113ΔCla, which represents the deletion of HIS4 and 3' AOX1, along with small stretches of pBR322 sequences used to link these sequences. The deletion of the HIS4, 3' AOX1 and pBR322 sequences removes one of two Csp45I sites present in the pHSA113 vector. The remaining Csp45I site is in the AOX1 5' regulatory region (promoter).

45 B. Creation of pXHSA113ΔCla

Digest 5 µg of pHSA113ΔCla for 1 hour at 37° C with 10 units of BstEII in 100 µl of REact 2 buffer. The digestion mixture was extracted with phenol and precipitated as detailed in step A. The DNA precipitate was dissolved in 100 µl of Csp45I buffer and digested at 37° C for 2 hours in the presence of 10 units of Csp45I. The digested DNA was then phenol extracted and precipitated as described in step A. The DNA precipitate was dissolved in 20 µl of water and 10 µl aliquots were loaded on 2 neighboring wells of a 0.9% agarose gel. Following electrophoresis, the gel portion corresponding to one of the lanes was stained and this was used to locate the position of the Csp45I-BstEII fragment of pHSA113ΔCla in the unstained lane. The gel portion containing the larger Csp45I-BstEII fragment was excised out and the DNA in the gel was

electroeluted into 500 μ l of 5 mM EDTA, pH 8.0. The DNA solution was phenol extracted as detailed in step A and the DNA precipitate was dissolved in 100 μ l water. The larger Csp45I-BstEII fragment was then ligated with the BstEII-Csp45I oligonucleotide linker described below. An aliquot (10 μ l) was ligated overnight at 4°C with 20 ng of annealed linker oligonucleotides 5'-CGAAACG ATG AAG TGG (SEQ ID NO:4) and 5'-GTTACCCACTTCATCGTTT (SEQ ID NO:5) in 20 μ l ligase buffer containing 100 μ g/ml BSA and 1 unit of T₄ DNA ligase. The ligation mixture was used to transform *E. coli* DG75' to obtain pXHSA113 Δ Cla. The pXHSA113 Δ Cla vector by virtue of the linker described above has an exact linkage between the 3' end of the native AOX1 5' regulatory region (promoter) and the HSA ATG start codon with no extraneous DNA sequences.

C. Creation of pHSA313

1 μ g of pXHSA113 Δ Cla was digested for 4 hours at 37°C with ClaI in 100 μ l of REact 1 buffer. Following digestion the reaction mixture was adjusted to alkaline phosphatase buffer conditions and treated with 10 units of calf intestinal alkaline phosphatase in a 200 μ l reaction volume for 30 minutes at 37°C. Phosphatase treatment was terminated by phenol extraction and the DNA was precipitated and dissolved in water at a concentration of approximately 10 ng/ μ l as described in step A and stored at -20°C.

1 μ g of pAO807N (Figure 8, construction of which is described in European Patent Application 0 344 459) was digested for 4 hours at 37°C with PstI in 100 μ l of REact 2 buffer. The digested DNA was adjusted to alkaline phosphatase buffer conditions and treated with 10 units of calf intestinal alkaline phosphatase in a 200 μ l reaction volume for 15 minutes at 55°C. At the end of 15 minutes another 10 units of phosphatase was added and incubated for 15 minutes. Phosphatase treatment was terminated by phenol extraction and the DNA was precipitated as described in step A. DNA was digested for 4 hours at 37°C with 5 units of ClaI in 100 μ l REact 1 buffer containing 100 μ g/ml BSA, followed by phenol extraction and precipitation of DNA as outlined in step A. The DNA precipitate was dissolved in water at a concentration of approximately 20 ng/ μ l.

Approximately 100 ng (10 μ l) of ClaI cleaved-phosphatased pXHSA113 Δ Cla was mixed with approximately 80 ng of PstI digested-phosphatased and ClaI-cleaved pAO807N (4 μ l), 4 μ l of 5X ligase buffer, 2 μ l of 1 mg/ml BSA and ligated overnight at 4°C using 1 unit of T₄ DNA ligase. The ligation mixture was used to transform *E. coli* DG75' to obtain pHSA313. The pHSA313 plasmid from this ligation contains the complete pXHSA113 Δ Cla sequence linked to the HIS4 gene and the AOX1 3' second insertable sequence derived from AO807N. The relative orientation of the components of the pHSA313 plasmid is the same as that shown in Figure 7 for plasmid pHSA113.

Example II

Construction of Expression Vector pPGP1

The expression vector pPGP1 was constructed in the following manner. pXHSA113 Δ Cla (see Example I) was digested with Bsu36I and PvuII (partial) and the vector backbone was isolated. An HSA structural gene on a PvuII-Bsu36I fragment analogous to the structural gene contained in pHSA113 (disclosed in European Patent Application 0 344 459) was ligated to this vector backbone to obtain pPGP1 Δ Cla. About 100 ng of pPGP1 Δ Cla was digested with ClaI at 37°C for 1 hour. The DNA was recovered as in Example I. About 100 ng of pAO807N (shown herein in Figure 8 and disclosed in European Patent Application 0 344 459) was digested with PstI, alkaline phosphatase treated and then digested with ClaI as detailed in Example I C. This fragment was then ligated to ClaI cleaned, alkaline phosphatase treated pPGP1 Δ Cla to obtain pPGP1. (GS115 pPGP1-9-6 is a clone which was obtained by transformation of *Pichia pastoris* GS115 with pPGP1 and this clone was used in fermentation).

Example III

Construction of 5' & 3' exact HSA expression plasmid pHSA413

The pHSA413 vector was constructed to provide a vector with an exact linkage between the 3' end of the AOX1 5' regulatory region and the start codon to the HSA structural gene as well as an exact linkage between the 5' end of the AOX1 3' termination sequence and the 3' end of the HSA structural gene.

A.. Creation of pXXHSA113 Δ Cla

1 µg of pXHSA113ΔCla was digested for 4 hours at 37°C with 10 units of EcoRI in 100 µl REact 3 buffer. The digestion mixture was phenol extracted and DNA precipitated as detailed in Example VI. DNA precipitate was dissolved in 20 µl water and digested for 1 hour at 37°C with 20 units of Bsu36I in 100 µl of Bsu36I buffer. The digestion mixture was phenol extracted, DNA precipitated and dissolved in 100 µl of water as detailed in Example VI. Approximately 100 ng of EcoRI and Bsu36I-cleaved DNA was mixed with 10 ng of annealed oligonucleotides 5'-TTAGGCTTATAAG (SEQ ID NO:6) and 5'-AATTCTTATAAGCC (SEQ ID NO:7) and ligated overnight at 4°C in 20 µl of T₄ DNA ligase buffer containing 100 µg/ml BSA and 10 units of T₄ DNA ligase. The ligation mixture was used to transform *E. coli* to obtain pXXHSA113ΔCla. In this plasmid the sequence between Bsu36I and EcoRI (SEQ ID NO:8) present in pXHSA113ΔCla shown below

Bsu36I

5' CCTTAGGCTTATAACATCTCTACATTTAAAAGCATCTCAGCCTACCATGAGAATAAGAGAAAGAAAATGAAGATCA
AAAGCTTATTTCATCTGTGTTTTCTTTTCGTTGGTGTAAGCCAACACCCTGTCTAAAAAACATAAATTTCTTTAATC
ATTTTGCCCTCTTTTCTCTGTGCTTCAATTAATAAAAAATGGAAGAATCTAAAAAAAAAAAAAAAAAGGAATTC
EcoRI

is replaced by 5' CC TTA GGC TTA TAA GAATTC (SEQ ID NO:9)

Bsu36I

EcoRI

B. Creation of pHSA413

1 µg of pXXHSA113ΔCla was digested for 4 hours at 37°C with ClaI in 100 µl of REact 1 buffer. Following digestion the reaction mixture was adjusted to alkaline phosphatase buffer conditions and treated with 10 units of calf intestinal alkaline phosphatase in 200 µl reaction volume for 30 minutes at 37°C. Phosphatase treatment was terminated by phenol extraction and the DNA was precipitated and dissolved in water at a concentration of approximately 10 ng/µl as described in step A and stored at -20°C.

Approximately 100 ng (10 µl) of ClaI cleaved-phosphatased pXXHSA113ΔCla was mixed with approximately 80 ng (4 µl) of PstI digested phosphatased and ClaI-cleaved pAO807N (see paragraph 2 in step 3 of Example VI), 4 µl of 5X ligase buffer, 2 µl of 1 mg/ml BSA and ligated overnight at 4°C using 1 unit of T₄ DNA ligase. The ligation mixture was used to transform *E. coli* DG75' to obtain pHSA413. The pHSA413 plasmid from this ligation contains the complete pXHSA113ΔCla sequence linked to the HIS4 gene and the AOX1 3' second insertable sequence derived from AO807N. The relative orientation of the components of the pHSA413 plasmid is the same as that shown in Figure 7 for plasmid pHSA113.

Example IV

Transformation of *Pichia pastoris* with pHSA313, pHSA413, and pPGP1

A.. Vector preparation

About 10 µg each of pHSA313, pHSA413, pPGP1, and pAO807N (negative control) were digested for 12 hours at 37°C in 200 µl of HS buffer with 50 units of NotI. The digested DNA samples were phenol extracted, precipitated as described in Example VI, dissolved in 20 µl of CaS, and were then used for transformation of *Pichia pastoris* GS115. About 10 µg each of pHSA313, pHSA413, and pAO807N were also digested with 20 units of SstI for 12 hours at 37°C in 200 µl of REact 2 buffer containing 100 µg/ml of BSA. The digested DNA samples were extracted with phenol, precipitated as described in Example VI and dissolved in 20 µl of CaS.

B. Cell Growth

Pichia pastoris GS115 (NRRL Y-15851) was inoculated into about 10 ml of YPD medium and shake cultured at 30°C for 12-20 hours. 100 ml of YPD medium was inoculated with a seed culture to give an OD₆₀₀ of about 0.001. The medium was cultured in a shake flask at 30°C for about 12-20 hours. The

culture was harvested when the OD₆₀₀ was about 0.2-0.3 by centrifugation at 1555 g for 5 minutes using a Sorvall RB5C.

C. Preparation of Spheroplasts

5 The cells were washed in 10 ml of sterile water, and then centrifuged at 1500 g for 5 minutes. (Centrifugation is performed after each cell wash at 1500 g for 5 minutes using a Sorvall RT6000B unless otherwise indicated.) The cells were washed once in 10 ml of freshly prepared SED, once in 10 ml of sterile 1M sorbitol, and finally resuspended in 10 ml of SCE buffer. 7.5 µl of 3 mg/ml Zymolyase (100,000 units/g, 10 obtained from Miles Laboratories) was added to the cell suspension. The cells were incubated at 30°C for about 10 minutes. (A reduction of 60% in OD₆₀₀ in 5% SDS can be utilized as a correct time marker.) The spheroplasts were washed in 10 ml of sterile 1 M sorbitol by centrifugation at 700 g for 5-10 minutes. 10 ml of sterile CaS was used as a final cell wash, and the cells were centrifuged again at 700 g for 5-10 minutes and then resuspended in 0.6 ml of CaS.

15 D. Transformation

Pichia pastoris GS115 cells were transformed with 10 µg of linearized DNA (see step A) using the spheroplast transformation technique of Sreekrishna et al, Gene 59, 115-125 (1987). DNA samples were 20 added (up to 20 µl volume) to 12 x 75 mm sterile polypropylene tubes. (DNA should be in a suitable buffer such as TE buffer or CaS.) 100 µl of spheroplasts were added to each DNA sample and incubated at room temperature for about 20 minutes. 1 ml of PEG solution was added to each sample and incubated at room temperature for about 15 minutes and centrifuged at 700 g for 5-10 minutes. SOS (150 µl) was added to the pellet and incubated for 30 minutes at room temperature. Finally 850 µl of 1M sorbitol was added.

25 E. Regeneration of Spheroplasts

A bottom agarose layer of 20 ml of regeneration agar SDR was poured per plate at least 30 minutes before transformation samples were ready. In addition, 8 ml aliquots of regeneration agar were distributed to 30 15 ml conical bottom Corning tubes in a 45°C water bath during the period that transformation samples were in SOS. Aliquots of 50 or 250 µl of the transformed sample was added to the 8 ml aliquots of molten regeneration agar held at 45°C and poured onto plates containing the solid 20 ml bottom agar layer. The plates were incubated at 30°C for 3-5 days.

35 F. Selection of Transformants

Transformants were selected for by culturing on SDR, a media lacking histidine. The colonies which grew in the absence of histidine were also screened for "methanol-slow" phenotype, indicating displacement of the AOX1 structural gene by the NotI DNA fragment) in the case of transformants obtained using 40 NotI linearized vectors. Several transformed GS115 cells showing "methanol-normal" (those obtained with SstI linearized DNA) and methanol-slow were then cultured and assayed for the production of HSA.

Example V

45 Methanol Induced Secretion of HSA in *Pichia pastoris* Integrative Transformants

Pichia pastoris GS115 strains transformed with pHSA313, pHSA413, and pPGP1 were analysed for HSA secretion in shake tube cultures. Both methanol-slow and methanol-normal strains were used. In each case 36 independent clones were studied. Transformants obtained with pAO807N served as negative 50 controls. A protocol was developed to ensure efficient secretion and stable accumulation of HSA in the culture medium.

Cells were grown to saturation in 10 ml BMGR or BMGY, and were placed in 50 ml tubes (2-3 days). The cells would be in the range of 10-20 A₆₀₀ units. The cells were harvested, the supernatant liquid was discarded, and then the pellet was resuspended in 2 ml of BMMR or BMMY. The tube was covered with a 55 sterile gauze (cheese cloth) instead of a cap. The tube(s) were then returned to a 30°C shaker. At the end of 2-3 days, the cells were pelleted, and the supernatant assayed for product. The pellets could be resuspended with fresh medium and returned to the shaker for renewed secretion. With *Pichia*-HSA strains, 10 µl of media supernatant was sufficient for analysis by SDS-PAGE followed by Coomassie staining. Under

these conditions a single band of 67 kD corresponding to HSA was observed. There was no significant difference between the expression levels of GS115/pHSA313 vs GS115/pHSA413 transformants, suggesting that deleting the 3' untranslated sequences from the HSA gene present in pHSA313 did not significantly affect expression levels. No significant difference in the HSA expression level was observed between methanol-slow vs methanol-normal transformants, suggesting that disruption of AOX1 was not essential for efficient HSA expression. As expected, HSA was absent in both the culture medium and the cell extract of GS115/pAO807N transformants (negative control). Clonal variants were selected which demonstrated increased levels of HSA secretion.

10 Example VI

Batch-Fed Fermentation of Mut⁻ *Pichia pastoris* for Production of HSA

Pichia pastoris GS115:pHSA 413-6 and pPGP1-9-6 were inoculated into two 20 liter BiolaFitte fermenters with an 8.5 l working volume. The inoculum was prepared in the following manner: a culture was grown on a YM plate and then transferred to 100 ml YM broth in a shake flask and grown for about 24 hours. 50 mls of this culture was transferred to 1 liter of YM broth in a shake flask and also grown for about 24 hours. 1 liter of this was then transferred to 8.5 liters of fermenter medium in the BiolaFitte fermenter. Fermentor medium consisted of Minimal salts + biotin + 5 percent glycerol. Batch growth conditions included the following: pH = 5.8 (controlled with NH₃), temperature = 30° C, and percent dissolved oxygen greater than 20 percent air saturation.

Glycerol exhaustion was complete after about 24 hours, at which time a slow methanol feed was begun at a rate of 10-15 ml/hr. The methanol concentration was monitored in the fermenter and the feed rate was adjusted to maintain a concentration of 0.5-0.9 percent of methanol in the broth.

Secreted HSA in the media was measured quantitatively by densitometry of Coomassie blue stained polyacrylamide gels containing SDS (SDS-PAGE). Areas were referenced to a series of known weights of authentic HSA run on the same SDS-PAGE gels. The data from these gels is included in Tables I and II.

The following Table illustrates the effect of changes in pH on the amount of HSA produced:

30 Table III

Production of HSA by Batch- Fed Fermentation			
Run	Strain	pH	HSA g/l
1	GS115:pPGP1-9-6	5.09-5.32	0.71
2	GS115:pPGP1-9-6	5.22	0.81
3	GS115:pPGP1-9-6	5.91	1.28
4	GS115:pPGP1-9-6	5.78	1.59
5	GS115:pPGP1-9-6	5.78	1.98
6	GS115:pPGP1-9-6	5.79	1.32

The following Table illustrates the level of HSA production which can be achieved at higher pH levels:

45 Table IV

Production of HSA by Batch-Fed Fermentation					
Run	Strain	pH	Hours MeOH	Dry Cell Wt.	HSA Broth g/l
1	GS115:pHSA 413-6	5.79	101	ND	2.13
2	GS115:pHSA 413-6	5.85	237	101	3.39
3	GS115:pHSA 413-6	5.85	265	98	2.70
4	GS115:pHSA 413-6	5.97	258	117	2.90
ND = Not Determined					

Example VII

Protocol for Shake Tube and Shake Flask Secretion of Proteins from *P. pastoris*

For efficient secretion and stable accumulation of HSA in shake tubes and shake flasks it is necessary to use a pH of 5.7-6.4 instead of 5.0 or 5.2 for the fermenter media, to add small amounts yeast extract (0.5-0.1%) and peptone (0.1-0.2%) to the fermenter medium and to start inducing expression at a low cell density (20-25 gram dry cell weight/liter). Using these techniques, we have developed a protocol that permits efficient secretion of HSA from cells grown in shake tubes and flasks. We believe that this protocol is applicable in general to secretion of proteins from *Pichia pastoris*.

10 Shake Tube:

Grow cells to saturation in 10 ml BMGR or BMGY placed in 50 ml tube (2-3 days). The A_{600} of cells will be in the range of 10-20. Harvest cells, discard the supernatant liquid and resuspend the pellet with 2 ml of BMMR or BMMY. Cover the tube with a sterile gauze or cheese cloth instead of the cap. Return the tube(s) to the shaker and maintain the shaker at about 30°C. At the end of 2-3 days, pellet cells, and analyze supernatant for product. The pellet can be resuspended with fresh media and returned to shaker for renewed secretion. With *Pichia*-HSA strains, 10 μ l of media supernatant is sufficient for analysis by SDS-PAGE followed by Coomassie staining. Under these conditions, a single band corresponding to HSA size (67 kD) is observed.

20

Shake Flask:

Grow cells as described above in 1 liter of medium (BMGY or BMGR) in a 2 liters flask. Harvest cells and suspend with 50-75 ml of BMMR or BMMY in a fermenter flask (Tunair™ shake-flask fermentation system, Research Products International Corporation) or a baffled flask covered with cheese cloth. Return to the shaker at 30°C and induce for 2-4 days. At the end of 2-4 days the cells are pelleted and the supernatant is analyzed for product. Shake tubes secretion can be re-initiated by resuspending the pelleted cells in fresh media.

30

35

40

45

50

55

SEQUENCE LISTING

5 (1) GENERAL INFORMATION:

(i) APPLICANT: William D. Prevatt et al.

10 (ii) TITLE OF INVENTION: Expression of Human Serum Albumin in
Pichia pastoris

(iii) NUMBER OF SEQUENCES: 3

15 (IV) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: RICHMOND, PHILLIPS, HITCHCOCK & UMPHLETT

(B) STREET: P.O. Box 2443

20 (C) CITY: Bartlesville

(D) STATE: OK.

(E) COUNTRY: USA

25 (F) ZIP: 74005

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Diskette

30 (B) COMPUTER: IBM PC

(C) OPERATING SYSTEM: PC-DOS/MS-DOS

(D) SOFTWARE: Display Write 4

35 (vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:

(B) FILING DATE:

40 (C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

45 (A) NAME: Hal Brent Woodrow

(B) REGISTRATION NUMBER: 32,501

(C) REFERENCE/DOCKET NUMBER: 32819US

50 (ix) TELECOMMUNICATION NUMBER: 1-918-661-0624

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 940 bp
 (B) TYPE: DNA
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(ix) SEQUENCE DESCRIPTION: SEQ ID NO:1:

5	AGATCTAACA	TCCAAAGACG	AAAGGTGAA	TGAAACCTTT	TGCCATCCG	ACATCCACAG	60
20	GTCCATTCTC	ACACATAAGT	GCCAAACGCA	ACAGGAGGGG	ATACACTAGC	AGCAGACCGT	120
	TGCAAACGCA	GGACCTCCAC	TCCTCTTCTC	CTCAACACCC	ACTTTTGCCA	TCGAAAAACC	180
	AGCCCAGTTA	TTGGGCTTGA	TTGGAGCTCG	CTCATTCCAA	TTCTTCTAT	TAGGCTACTA	240
25	ACACCATGAC	TTTATTAGCC	TGTCIATCCT	GGCCCCCCTG	GCGAGGTTC	TGTTTGTTTA	300
	TTTCCGAATG	CAACAAGCTC	CGCATTACAC	CCGAACATCA	CTCCAGATGA	GGGCTTTCTG	360
	AGTGTGGGGT	CAAATAGTTT	CATGTTCCCC	AAATGGCCCA	AAACTGACAG	TTTAAACGCT	420
30	GTCTTGGAAC	CTAATATGAC	AAAAGCGTGA	TCTCATCCAA	GATGAACTAA	GTTTGGTTCTG	480
	TTGAAATGCT	AACGGCCAGT	TGGTCAAAAA	GAAACTTCCA	AAAGTCGGCA	TACCGTTTGT	540
35	CTTGTTTGGT	ATTGATTGAC	GAATGCTCAA	AAATAATCTC	ATTAATGCTT	AGCGCAGTCT	600
	CTCTATCGCT	TCTGAACCCC	GGTGCACCTG	TGCCGAAACG	CAAATGGGGA	AACACCCGCT	660
	TTTTGGATGA	TTATGCATTG	TCTCCACATT	GTATGCTTCC	AAGATTCTGG	TGGGAATACT	720
40	GCTGATAGCC	TAACGTTTAT	GATCAAAATT	TAAGTGTCT	AACCCCTACT	TGACAGCAAT	780
	ATATAAACAG	AAGGAAGCTG	CCCTGTCTTA	AACCTTTTTT	TTTATCATCA	TTATTAGCTT	840
	ACTTTCATAA	TTGCGACTGG	TTCCAATTGA	CAAGCTTTTG	ATTTTAACGA	CTTTTAACGA	900
45	CAACTTGAGA	AGATCAAAAA	ACAACTAATT	ATTCGAAACG			940

(3) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 600 bp
 (B) TYPE: DNA
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(ix) SEQUENCE DESCRIPTION: SEQ ID NO:2:

5	AAAGTAAACC	CCATTCAATG	TTCCGAGATT	TAGTATACTT	GCCCCTATAA	GAAACGAAGG	60
15	ATTTTCAGCTT	CCTTACCCCA	TGAACAGAAA	TCTTCCATTT	ACCCCCCACT	GGAGAGATCC	120
	GCCCAAACGA	ACAGATAATA	GAAAAAAGAA	ATTCGGACAA	ATAGAACACT	TTCTCAGCCA	180
	ATTAAAGTCA	TTCCATGCAC	TCCCTTTAGC	TGCCGTTCCA	TCCCTTTGTT	GAGCAACACC	240
20	ATCGTTAGCC	AGTACGAAAG	AGGAAACTTA	ACCGATACCT	TGGAGAAATC	TAAGGCGCGA	300
	ATGAGTTTAG	CCTAGATATC	CTTAGTGAAG	GGTGTTCGA	TACCTTCTCC	ACATTCAGTC	360
	ATAGATGGGC	AGCTTTGTIA	TCATGAAGAG	ACGGAAACGG	GCATTAAGGG	TTAACCGCCA	420
25	AATTATATAA	AAGACAACAT	GTCCCCAGTT	TAAAGTTTTT	CTTTCCTATT	CTTGTATCCT	480
	GAGTGACCGT	TGTGTTTAAT	ATAACAAGTT	CGTTTTAACT	TAAGACCAAA	ACCAGTTACA	540
30	ACAAATTATA	ACCCCTCTAA	ACACTAAAGT	TCACTCTTAT	CAAACATCA	AACATCAAAA	600

(4) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1830 bp
 (B) TYPE: DNA
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(ix) SEQUENCE DESCRIPTION: SEQ ID NO:3:

EP 0 510 678 A2

	ATG Met	AAG Lys -35	TGG Trp	GTA Val	ACC Thr	TTT Phe	ATT Ile -30	TCC Ser	CTT Leu	CTT Leu	TTT Phe	CTC Leu -25	TTT Phe	AGC Ser	TCG Ser
5	GCT Ala	TAT Tyr -20	TCC Ser	AGG Arg	GGT Gly	GTG Val	TTT Phe -15	CGT Arg	CGA Arg	GAT Asp	GCA Ala	CAC His -10	AAG Lys	AGT Ser	GAG Glu
10	GTT Val	GCT Ala -5	CAT His	CGG Arg	TTT Phe	AAA Lys	GAT Asp 1	TTG Leu	GGA Gly	GAA Glu	GAA Glu 5	AAT Asn	TTC Phe	AAA Lys	GCC Ala
15	TTG Leu 10	GTG Val	TTG Leu	ATT Ile	GCC Ala	TTT Phe 15	GCT Ala	CAG Gln	TAT Tyr	CTT Leu	CAG Gln 20	CAG Gln	TGT Cys	CCA Pro	TTT Phe
	GAA Glu 25	GAT Asp	CAT His	GTA Val	AAA Lys	TTA Leu 30	GTG Val	AAT Asn	GAA Glu	GTA Val	ACT Thr 35	GAA Glu	TTT Phe	GCA Ala	AAA Lys
20	ACA Thr 40	TGT Cys	GTT Val	GCT Ala	GAT Asp	GAG Glu 45	TCA Ser	GCT Ala	GAA Glu	AAT Asn	TGT Cys 50	GAC Asp	AAA Lys	TCA Ser	CTT Lue
25	CAT His 55	ACC Thr	CTT Leu	TTT Phe	GGA Gly	GAC Asp 60	AAA Lys	TTA Leu	TGC Cys	ACA Thr	GTT Val 65	GCA Ala	ACT Thr	CTT Leu	CGT Arg
	GAA Glu 70	ACC Thr	TAT Tyr	GGT Gly	GAA Glu	ATG Met 75	GCT Ala	GAC Asp	TGC Cys	TGT Cys	GCA Ala 80	AAA Lys	CAA Gln	GAA Glu	CCT Pro
30	GAG Glu 85	AGA Arg	AAT Asn	GAA Glu	TGC Cys	TTC Phe 90	TTG Leu	CAA Gln	CAC His	AAA Lys	GAT Asp 95	GAC Asp	AAC Asn	CCA Pro	AAC Asn
35	CTC Leu 100	CCC Pro	CGA Arg	TTG Leu	GTG Val	AGA Arg 105	CCA Pro	GAG Glu	GTT Val	GAT Asp	GTG Val 110	ATG Met	TGC Cys	ACT Thr	GCT Ala
40	TTT Phe 115	CAT His	GAC Asp	AAT Asn	GAA Glu	GAG Glu 120	ACA Thr	TTT Phe	TTG Leu	AAA Lys	AAA Lys 125	TAC Tyr	TTA Leu	TAT Tyr	GAA Glu
45															
50															
55															

5	ATT Ile 130	GCC Ala	AGA Arg	AGA Arg	CAT His	CCT Pro 135	TAC Tyr	TTT Phe	TAT Tyr	GCC Ala	CCG Pro 140	GAA Glu	CTC Leu	CTT Leu	TTC Phe
	TTT Phe 145	GCT Ala	AAA Lys	AGG Arg	TAT Tyr	AAA Lys 150	GCT Ala	GCT Ala	TTT Phe	ACA Thr	GAA Glu 155	TGT Cys	TGC Cys	CAA Gln	GCT Ala
10	GCT Ala 160	GAT Asp	AAA Lys	GCT Ala	GCC Ala	TGC Cys 165	CTG Leu	TTG Leu	CCA Pro	AAG Lys	CTC Leu 170	GAT Asp	GAA Glu	CTT Leu	CGG Arg
15	GAT Asp 175	GAA Glu	GGG Gly	AAG Lys	GTT Val	TCG Ser 180	TCT Ser	GCC Ala	AAA Lys	CAG Gln	AGA Arg 185	CTC Leu	AAG Lys	TGT Cys	GCC Ala
	AGT Ser 190	CTC Leu	CAA Gln	AAA Lys	TTT Phe	GGA Gly 195	GAA Glu	AGA Arg	GCT Ala	TTC Phe	AAA Lys 200	GCA Ala	TGG Trp	GCA Ala	GTA Val
20	GCT Ala 205	CGC Arg	CTG Leu	AGC Ser	CAG Gln	AGA Arg 210	TTT Phe	CCC Pro	AAA Lys	GCT Ala	GAG Glu 215	TTT Phe	GCA Ala	GAA Glu	GTT Val
25	TCC Ser 220	AAG Lys	TTA Leu	GTG Val	ACA Thr	GAT Asp 225	CTT Leu	ACC Thr	AAA Lys	GTC Val	CAC His 230	ACG Thr	GAA Glu	TGC Cys	TGC Cys
30	CAT His 235	GGA Gly	GAT Asp	CTG Leu	CTT Leu	GAA Glu 240	TGT Cys	GCT Ala	GAT Asp	GAC Asp	AGG Arg 245	GCG Ala	GAC Asp	CTT Leu	GCC Ala
	AAG Lys 250	TAT Tyr	ATC Ile	TGT Cys	GAA Glu	AAT Asn 255	CAA Gln	GAT Asp	TCG Ser	ATC Ile	TCC Ser 260	AGT Ser	AAA Lys	CTG Leu	AAG Lys
35	GAA Glu 265	TGC Cys	TGT Cys	GAA Glu	AAA Lys	CCT Pro 270	CTG Leu	TTG Leu	GAA Glu	AAA Lys	TCC Ser 275	CAC His	TGC Cys	ATT Ile	GCC Ala
40	GAA Glu 280	GTG Val	GAA Glu	AAT Asn	GAT Asp	GAG Glu 285	ATG Met	CCT Pro	GCT Ala	GAC Asp	TTG Leu 290	CCT Pro	TCA Ser	TTA Leu	GCT Ala
	GCT Ala 295	GAT Asp	TTT Phe	GTT Val	GAA Glu	AGT Ser 300	AAG Lys	GAT Asp	GTT Val	TGC Cys	AAA Lys 305	AAC Asn	TAT Tyr	GCT Ala	GAG Glu
45	GCA Ala 310	AAG Lys	GAT Asp	GTC Val	TTC Phe	TTG Leu 315	GGC Gly	ATG Met	TTT Phe	TTG Leu	TAT Tyr 320	GAA Glu	TAT Tyr	GCA Ala	AGA Arg
50															
55															

EP 0 510 678 A2

	AGG Arg 325	CAT His	CCT Pro	GAT Asp	TAC Tyr	TCT Ser 330	GTC Val	GTG Val	CTG Leu	CTG Leu	CTG Leu 335	AGA Arg	CTT Leu	GCC Ala	AAG Lys
5	ACA Thr 340	TAT Tyr	GAA Glu	ACC Thr	ACT Thr	CTA Leu 345	GAG Glu	AAG Lys	TGC Cys	TGT Cys	GCC Ala 350	GCT Ala	GCA Ala	GAT Asp	CCT Pro
10	CAT His 355	GAA Glu	TGC Cys	TAT Tyr	GCC Ala	AAA Lys 360	GTG Val	TTC Phe	GAT Asp	GAA Glu	TTT Phe 365	AAA Lys	CCT Pro	CTT Leu	GTG Val
15	GAA Glu 370	GAG Glu	CCT Pro	CAG Gln	AAT Asn	TTA Leu 375	ATC Ile	AAA Lys	CAA Gln	AAT Asn	TGT Cys 380	GAG Glu	CTT Leu	TTT Phe	GAG Glu
20	CAG Gln 385	CTT Leu	GGA Gly	GAG Glu	TAC Tyr	AAA Lys 390	TTC Phe	CAG Gln	AAT Asn	GCG Ala	CTA Leu 395	TTA Leu	GTT Val	CGT Arg	TAC Tyr
25	ACC Thr 400	AAG Lys	AAA Lys	GTA Val	CCC Pro	CAA Gln 405	GTG Val	TCA Ser	ACT Thr	CCA Pro	ACT Thr 410	CTT Leu	GTA Val	GAG Glu	GTC Val
30	TCA Ser 415	AGA Arg	AAC Asn	CTA Leu	GGA Gly	AAA Lys 420	GTG Val	GGC Gly	AGC Ser	AAA Lys	TGT Cys 425	TGT Cys	AAA Lys	CAT His	CCT Pro
35	GAA Glu 430	GCA Ala	AAA Lys	AGA Arg	ATG Met	CCC Pro 435	TGT Cys	GCA Ala	GAA Glu	GAC Asp	TAT Tyr 440	CTA Leu	TCC Ser	GTG Val	GTC Val
40	CTG Leu 445	AAC Asn	CAG Gln	TTA Leu	TGT Cys	GTG Val 450	TTG Leu	CAT His	GAG Glu	AAA Lys	ACG Thr 455	CCA Pro	GTA Val	AGT Ser	GAC Asp
45	AGA Arg 460	GTC Val	ACC Thr	AAA Lys	TGC Cys	TGC Cys 465	ACA Thr	GAA Glu	TCC Ser	TTG Leu	GTG Val 470	AAC Asn	AGG Arg	CGA Arg	CCA Pro
50	TGC Cys 475	TTT Phe	ICA Ser	GCT Ala	CTG Leu	GAA Glu 480	GTC Val	GAT Asp	GAA Glu	ACA Thr	TAC Tyr 485	GTT Val	CCC Pro	AAA Lys	GAG Glu
55	TTT Phe 490	AAT Asn	GCT Ala	GAA Glu	ACA Thr	TTC Phe 495	ACC Thr	TTC Phe	CAT His	GCA Ala	GAT Asp 500	ATA Ile	TGC Cys	ACA Thr	CTT Leu
	TCT Ser 505	GAG Glu	AAG Lys	GAG Glu	AGA Arg	CAA Gln 510	ATC Ile	AAG Lys	AAA Lys	CAA Gln	ACT Thr 515	GCA Ala	CTT Leu	GTT Val	GAG Glu

5 CTT GTG AAA CAC AAG CCC AAG GCA ACA AAA GAG CAA CTG AAA GCT
 Leu Val Lys His Lys Pro Lys Ala Thr Lys Glu Gln Leu Lys Ala
 520 525 530
 GTT ATG GAT GAT TTC GCA GCT TTT GTA GAG AAG TGC TGC AAG GCT
 Val Met Asp Asp Phe Ala Ala Phe Val Glu Lys Cys Cys Lys Ala
 535 540 545
 10 GAC GAT AAG GAG ACC TGC TTT GCC GAG GAG GGT AAA AAA CTT GTT
 Asp Asp Lys Glu Thr Cys Phe Ala Glu Glu Gly Lys Lys Leu Val
 550 555 560
 GCT GCA AGT CAA GCT GCC TTA GGC TTA TAA
 Ala Ala Ser Gln Ala Ala Leu Gly Leu -
 15 565 570

(5) INFORMATION FOR SEQ ID NO:4:

20 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 16bp
 (B) TYPE: DNA
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: Oligonucleotide
 25 (ix) SEQUENCE DESCRIPTION: SEQ ID NO:4:

30 CGAAACG ATG AAG TGG 16
 Met Lys Trp

(6) INFORMATION FOR SEQ ID NO:5:

35 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 19bp
 (B) TYPE: DNA
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: Oligonucleotide
 40 (ix) SEQUENCE DESCRIPTION: SEQ ID NO:5:

45 GTTACCCACT TCATCGTTT 19

(7) INFORMATION FOR SEQ ID NO:6:

50 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 13bp
 (B) TYPE: DNA

55

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: Oligonucleotide

(ix) SEQUENCE DESCRIPTION: SEQ ID NO:6:

10 TTAGGCTTAT AAG 13

(8) INFORMATION FOR SEQ ID NO:7:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 14bp
(B) TYPE: DNA
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: Oligonucleotide

(ix) SEQUENCE DESCRIPTION: SEQ ID NO:7:

25 AATTCTTATA AGCC 14

(9) INFORMATION FOR SEQ ID NO:8:

30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 231bp
(B) TYPE: DNA
(C) STRANDED: single
(D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: Linker Oligonucleotide

(ix) SEQUENCE DESCRIPTION: SEQ ID NO:8:

40 CCTTAGGCTT ATAACATCTC TACATTTAAA AGCATCTCAG CCTACCATGA GAATAAGAGA 60
AAGAAAATGA AGATCAAAAG CTTATTCATC TGTGTTTTCT TTTTCGTTGG TGTAAGCCA 120
ACACCCTGTC TAAAAACAT AAATTTCTTT AATCATTTTG CCTCTTTTTC TCTGTGCTTC 180
45 AATTAATAAA AAATGGAAAG AATCTAAAAA AAAAAAAAAA AAAAGGAATT C 231

50

55

(10) INFORMATION FOR SEQ ID NO:9:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20bp
- (B) TYPE: DNA
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: Oligonucleotide

(1x) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CCTTAGGCTT ATAAGAATTC 20

Claims

1. An improved process for the secretion of a heterologous protein in transformed *Pichia pastoris* cells comprising:
 - (a) cultivating in a fermentation broth transformed *Pichia pastoris* cells capable of expressing a heterologous structural gene encoding a secretion signal sequence and a mature heterologous protein, wherein the signal sequence is operably linked to the sequence encoding the mature heterologous protein, under condition suitable for sustaining the viability of said transformed *Pichia pastoris* cells, under suitable conditions for the expression of said heterologous protein by said *Pichia pastoris* cells, and
 - (b) maintaining the pH of said fermentation broth at a pH in the range of 5.7 to 6.4 contemporaneously with the expression of a heterologous protein.
2. The process of claim 1 wherein the transformed *Pichia pastoris* cells are grown in a batch-fed manner during the expression of HSA and/or the pH of the fermentation broth is maintained during expression of the heterologous protein in the range of pH 5.7 to pH 6.0, preferably at a pH of 5.8.
3. The process of claim 2 wherein the fermentation broth contains an effective amount of a suitable minimal salts mixture, growth factors and at least one suitable carbon source selected from methanol, glycerol, sorbitol, glucose, fructose and combinations of two or more thereof to maintain the viability of said transformed *Pichia pastoris* cells.
4. The process of claim 3, wherein after the fermentation broth's carbon source is consumed, the transformed *Pichia pastoris* cells are contacted with methanol wherein the methanol is provided at a rate sufficient to maintain the viability of the *Pichia pastoris* cells in contact therewith and the methanol concentration dose not exceed about 5.0 percent by weight.
5. All improved process for the expression of HSA in transformed *Pichia pastoris* cells comprising:
 - (a) cultivating in a fermentation broth transformed *Pichia pastoris* cells capable of expressing HSA under conditions suitably for maintaining the viability of said transformed *Pichia pastoris* cells, under suitable conditions for the expression of HSA by said *Pichia pastoris* cells, and
 - (b) contacting said fermentation broth containing the transformed *Pichia pastoris* cells with a suitable amount of added amino acids and peptone to enhance the secretion of HSA contemporaneously with the expression of HSA and,
 - (c) optionally, maintaining the pH of the fermentation broth during expression of the heterologous protein in the range of from 5.7 to 6.0.
6. The process of claim 1 or 5 wherein *Pichia pastoris* is transformed with a vector selected from a circular plasmid and a linear plasmid, the latter preferably being a linear integrative site-specific vector.

7. The process of claim 6 wherein said linear integrative site-specific vector contains the following serial arrangement:
 - (a) a first insertable DNA fragment,
 - (b) at least one marker gene, and at least one expression cassette containing a heterologous structural gene encoding a signal sequence and a mature heterologous protein, operably linked to a regulatory region and a 3' termination sequence, and
 - (c) a second insertable DNA fragment
 wherein the order of the marker gene and cassette of component (b) may be interchanged, and the first and second insertable DNA fragments employed are homologous with separate portions of the *Pichia pastoris* genome wherein the insertable fragments are in the same relative orientation as exist in the *Pichia pastoris* genome.
8. The process of claim 7, wherein the first insertable DNA fragment and the second insertable DNA fragment are obtained from the DNA sequence of a gene from *Pichia pastoris* selected from the AOX1 gene, the p40 gene, the DAS gene, the GAP gene, the PHO1 gene and the HIS4 gene.
9. The process of claim 7 wherein said expression cassette comprises:
 - (a) a regulatory region selected from the AOX1 5' regulatory region isolated from *Pichia pastoris*, the p40 5' regulatory region isolated from *Pichia pastoris*, the DAS 5' regulatory region from *Pichia pastoris*, the GAP 5' regulatory region isolated from *Pichia pastoris*, the PHO1 5' regulatory region isolated from *Pichia pastoris*, the acid phosphatase promoter isolated from *Saccharomyces cerevisiae*, the galactosidase promoter isolated from *Saccharomyces cerevisiae*, the alcohol dehydrogenase promoter isolated from *Saccharomyces cerevisiae*, the alpha-mating factor promoter isolated from *Saccharomyces cerevisiae* and the glyceraldehyde 3-phosphate dehydrogenase promoter isolated from *Saccharomyces cerevisiae* operably linked to
 - (b) a heterologous structural gene encoding a secretion signal sequence and a mature heterologous protein, wherein the mature heterologous protein is selected from the group consisting of tissue plasminogen activator, albumins, lysozyme, interferon and invertase and the secretion signal is selected from the group consisting of a native signal sequence for the heterologous protein, a signal sequence of the *Pichia pastoris* acid phosphatase gene and a signal sequence of the *Saccharomyces cerevisiae* alpha-mating factor gene, and
 - (c) a 3' termination sequence from *Pichia pastoris* selected from the 3' termination sequence isolated from the AOX1 gene, the p40 gene, the DAS gene, the GAP gene, the PHO1 gene and the HIS4 gene.
10. The process of claim 7, wherein said marker gene is selected from HIS4 isolated from *Pichia pastoris*, ARG4 isolated from *Pichia pastoris*, SUC2 isolated from *Saccharomyces cerevisiae*, G418^R gene of Tn903 and G418^R gene of Tn601.
11. The process of claim 9, wherein the heterologous structural gene encodes the HSA native signal sequence operably linked to the sequence encoding the mature HSA protein.
12. The process of claim 9 wherein the plasmid comprises an autonomously replicating DNA sequence and a marker gene, said marker gene being selected from HIS4 isolated from *Pichia pastoris*, ARG4 isolated from *Pichia pastoris*, SUC2 isolated from *Saccharomyces cerevisiae*, G418^R gene of Tn903 and G418^R gene of Tn601.
13. The process of claim 12 wherein said plasmid comprises:
 - (a) the AOX1 5' regulatory region isolated from *Pichia pastoris* operably linked to
 - (b) a structural gene for HSA encoding a native signal sequence for HSA and a mature HSA protein, wherein the HSA signal sequence is operably linked to the sequence encoding the mature HSA protein operably linked to
 - (c) the 3' termination sequence of AOX1 isolated from *Pichia pastoris* operably linked to
 - (d) at least one marker gene, preferably a HIS4 gene, and
 - (e) a second DNA fragment which is about a 0.19 kilobase sequence of an autonomously replicating DNA sequence.
14. The process of claim 5 wherein the amino acids are provided in the form of yeast extract at a

EP 0 510 678 A2

concentration in the range of 1 g/liter to 15 g/liter and/or wherein the peptone is provided at a concentration in the range of 1 g/liter to 50 g/liter.

5

10

15

20

25

30

35

40

45

50

55

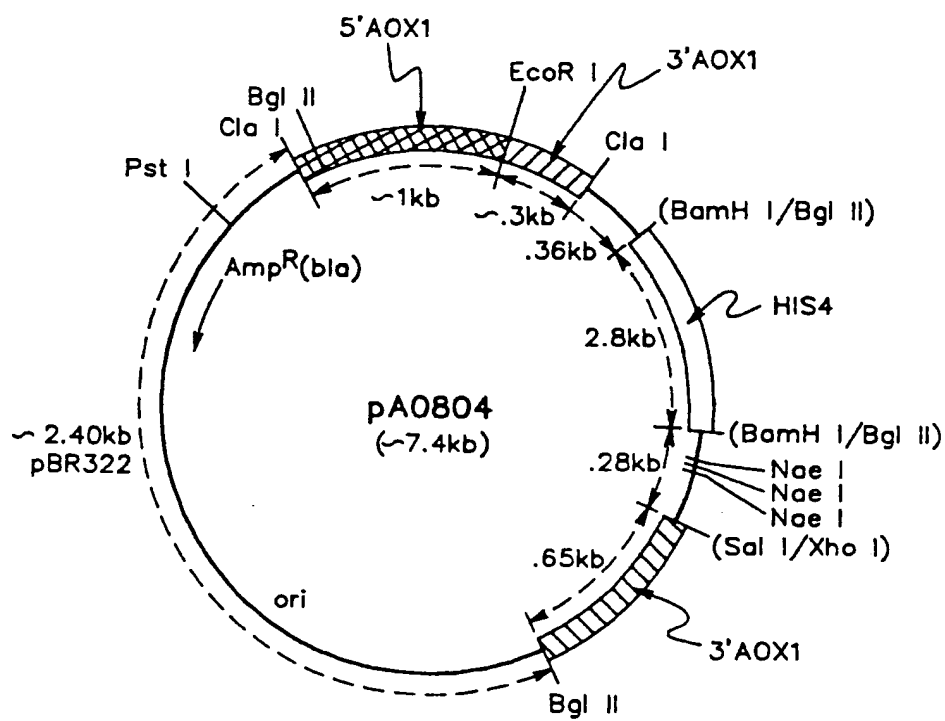


FIG. 1

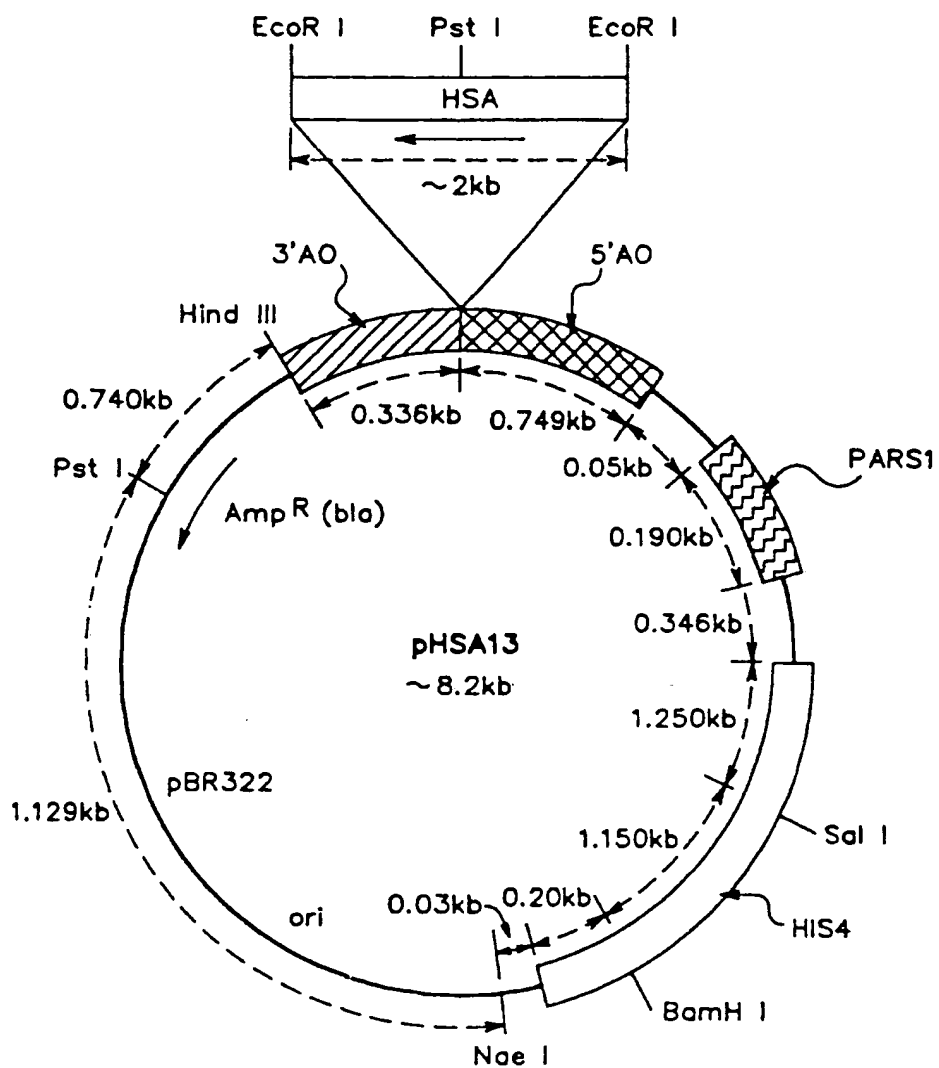


FIG. 2

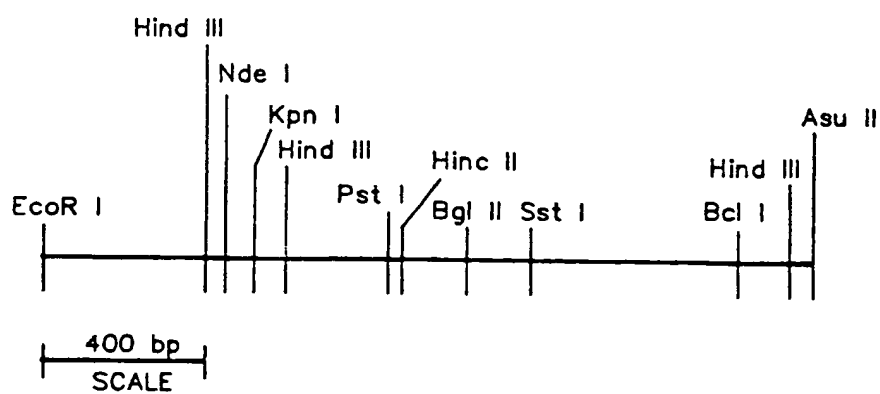


FIG. 3

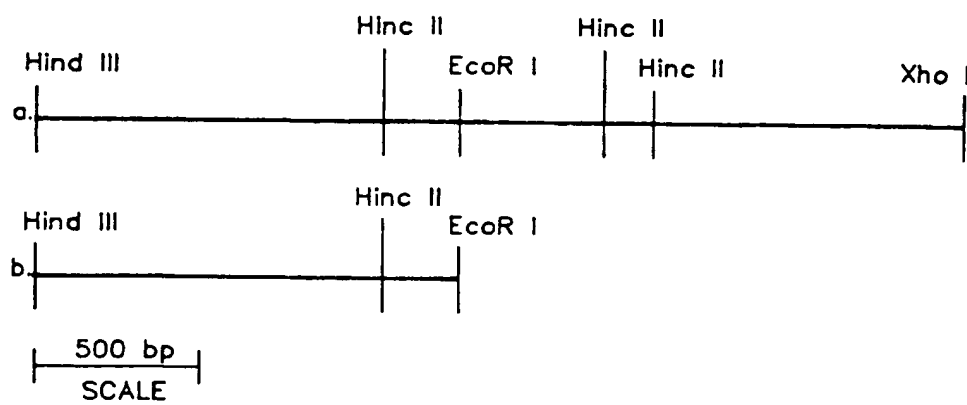


FIG. 4

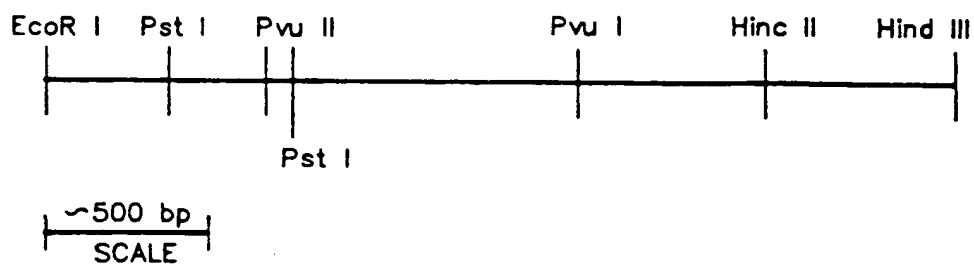


FIG. 5

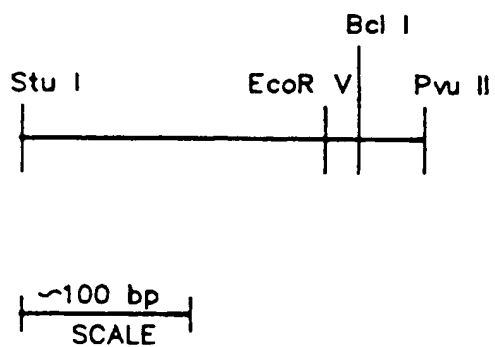
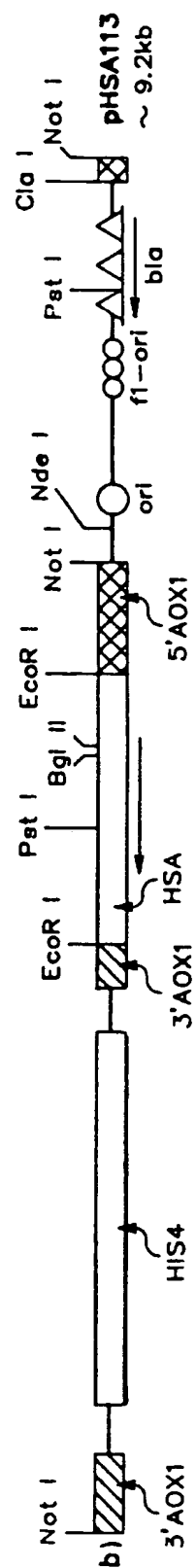


FIG. 6



SCALE: 1kb

FIG. 7

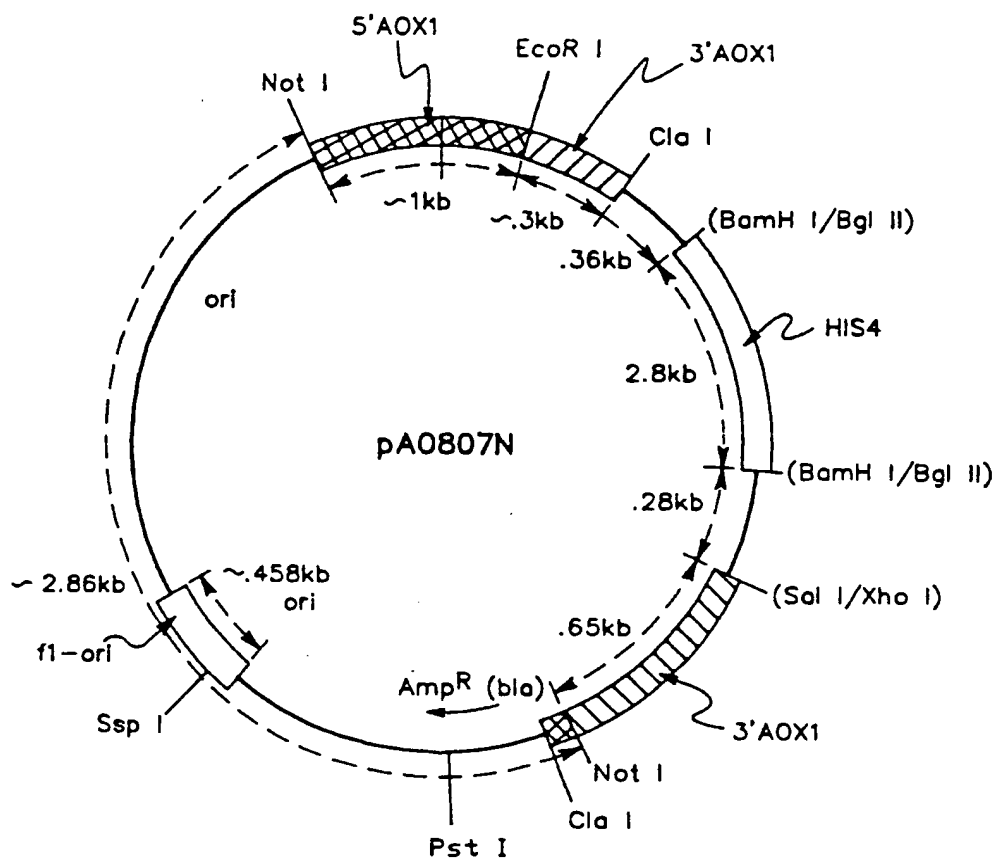


FIG. 8